DETERMINATION OF PROTONATION CONSTANTS FOR 2-HYDROXY-1-(2-HYDROXY-4-SULPHO-1-NAPHTHYLAZO)-3-NAPHTHOIC ACID

J.A. LIYANAGE* AND T. K. JANARATNE

Department of Chemistry, University of Kelaniya, Kelaniya, Sri Lanka

* Corresponding author (e-mail:janitha@kln.ac.lk)

ABSTRACT

Chemical speciation modelling in combination with potentiometric titration method was used to determine the protonation constants for 2-hydroxy-1-(2-hydroxy-4-sulpho-1-naphthylazo)-3-naphthoic acid, the Patton and Reeder's reagent, at 25° C in three ionic strengths of 0.1 mol dm⁻³, 0.01 mol dm⁻³ and 0.001 mol dm⁻³ NaNO₃. Potentiometric titrations were performed and pH, titrant volume, temperature and free and total proton concentrations at each titration point were measured. Constants were determined for three of the four protonated sites and the forth one being a very low value as this site is always in the ionic form and is readily ionized in aqueous solutions.

Keywords: Patton and Reeder, Protonation constants, Potentiometric titrations, Chemical speciation modelling

INTRODUCTION

Chemical speciation modelling has developed in recent years as an important tool for analytical chemists. It has been widely used to explain the behavior of metal ions in solutions (Duffield & Williams, 1989, Liyanage & Janaratna, 2002, Liyanage & Allas, 1999). In a similar fashion chemical speciation has played a critical role in the determination, correction and confirmation of thermodynamic formation constants

(Jackson & Seymour 1995; Jackson & Seymour 1995; De Robertis & et al., 1995, L-Gonzalez and et al., 1997).

2-hydroxy-1-(2-hydroxy-4-sulpho-1-naphthylazo)-3-naphthoic acid (Figure 1) which is named as Patton and Reeder's reagent and abbreviated as HHSNNA (H₄L) is mainly used as the indicator in the direct titration of calcium, particularly in the presence of magnesium, with ethylenediaminetetraacetic acid (EDTA).

Figure 1. 2-hydroxy-1-(2-hydroxy-4-sulpho-1-naphthylazo)-3-naphthoic acid (Patton and Reeder's Reagent)

The protonation constants of this compound are needed when these titrations are modeled to study the underlying chemical principles. However, protonation constants of the Patton and Reeder's reagent were not determined previously and were not found in well-established databases (Smith & Martell, 1989). Hence the overall aim of this investigation was to determine these constants using a potentiometric titration method in combination with chemical speciation modelling.

EXPERIMENTAL

Materials

Patton and Reeder's reagent used for the study was obtained from Loba chemie (India) and unless otherwise mentioned, all other chemicals were from BDH (UK) and Sigma (USA). Prior to distillation, water was passed through a mixed bed ion exchange resin to remove any ionic contaminants. Distilled water thus prepared was boiled for two hours to minimize atmospheric carbon dioxide contamination; there after it was cooled to ambient room temperature, 25° C, in a closed vessel leaving no headspace.

All titrations were conducted at 25° C under high purity (99.996%) nitrogen or argon-purged conditions.

Methods

Potentiometric titrations were performed to quantify protonation binding of Patton and Reeder's reagent using an auto-titrator/IBM PC data collection system (Orion Research Institute, USA, 960 auto chemistry analyzer) at 25° C in a glove box under nitrogen atmosphere. The glove box was constructed to minimize CO₂ contamination from atmospheric environments. The reaction vessel was placed in the glove box that is connected to the auto-titrator to monitor pH, titrant volume and temperature at each titration point. Free and total proton concentrations measured at 50 ul volume increments were captured by an IBM PC linked via RS232C interface (the minimum dispensing volume of the Orion 960 auto chemistry analyzer was 50 µl). Titration experiments were performed in three ionic strengths that are 0.1 mol dm⁻³, 0.01 mol dm⁻³ and 0.001 mol dm⁻³ NaNO₃ concentrations. Reagent interferences were minimized by performing blank titrations in all three ionic strengths. The volume versus pH data was interpolated by cubic spline technique to subtract the effects caused by the solution matrix. 10 ml aliquots of the titrant were placed in the glove box in tightly capped 25 ml polypropylene centrifuge tubes that were filled with N₂ previously. All samples were equilibrated in a water bath shaker at regulated temperature conditions at 25° C.

All pH measurements were made using Ross combination pH electrode and auto chemistry analyzer in well-stirred solutions.

The proton binding data was modeled using 1-pK basic Stern layer model (BSM). All calculations were carried out using FITEQL (Herbeli & Westll, 1994) and ECOSAT-FIT (Keizer & van Riemsdijk, 1999, Kinnburgh, 1993) algorithms.

RESULTS

Titrations for determination of protonation constants at three different ionic strengths, at 0.1 mol dm⁻³, 0.01 mol dm⁻³ and 0.001 mol dm⁻³ NaNO₃ were carried out

and the results are shown in the Table 1. The total concentration of the legend (H_4L) used was $0.01 mol\ dm^{-3}$.

Table 1. Protonation Constants of the Legend

Protonation Cons	tant	Ionic Strength \pm SD (n=10)				
		0.1 mol dm ⁻³	0.01 mol dm ⁻³	0.001 mol dm ⁻³		
$-$ Log β_{HL}	$H + L^{4-} = HL^{3-}$	10.550 ± 0.006	10.500 ± 0.004	10.600± 0.005		
$\log \beta_{\mathrm{H}_2\mathrm{L}}$	$2H + L^{4-} = H_2L^{2-}$	14.911 ± 0.005	15.352 ± 0.003	15.149 ± 0.003		
$\log \beta_{\mathrm{H_3L}}$	$3H + L^{4-} = H_3L^{-}$	17.298 ± 0.003	18.119 ± 0.004	17.895 ± 0.002		

The β values for the protonation of 2-hydroxy-1-(2-hydroxy-4-sulpho-1-naphthylazo)-3-naphthoic acid correspond to the stepwise protonation constants in the Table 2.

Table 2. Stepwise Protonation Constants

Stepwise Protonation Constant			0.1 mol dm ⁻³	Ionic Strength 0.01 mol dm ⁻³	0.001 mol dm ⁻³
$Log K_{HL}$	H + L ⁴⁻	= HL ³⁻	10.550 ± 0.006	10.500 ± 0.004	10.600± 0.005
$\text{Log}K_{\text{H}_2\text{L}}$	$H + HL^{3-}$	$= H_2L^{2-}$	4.361 ± 0.005	4.852 ± 0.003	4.549 ± 0.003
$\log K_{\mathrm{H_3L}}$	$H + H_2L^{2-}$	$= H_3L^-$	2.387 ± 0.003	2.767 ± 0.004	2.746 ± 0.002

DISCUSSION

The pK of only three of the four protons could be determined for this compound and the pK of the other proton being very low.

The high value of $\log K_{\rm HL}$ and the large difference between $\log K_{\rm HL}$ and $\log K_{\rm H_2L}$ may be indicative of the intramolecular hydrogen bonding between the protonated and deprotonated hydroxyl groups.

Three proton constants for 2-hydroxy-1-(2-hydroxy-4-sulpho-1-naphthylazo)-3-naphthoic acid could be determined in three different ionic strengths at 0.1 mol dm⁻³, 0.01 mol dm⁻³ and 0.001 mol dm⁻³ NaNO₃ media. This forth site is always in the ionic form and this site is readily ionized in aqueous solutions.

ACKNOWLEDGEMENTS

Authors acknowledge the Chemical Modeling Laboratory, Institute of Fundamental Studies, Kandy, Sri Lanka for providing facilities to use the Orion 960 auto-chemistry analyzer and the University of Kelaniya (grant No RP/RG/98/01/03) for financial support.

REFERENCES

Duffield, J. R. & D.R. Williams 1989. Chemistry in Britain April; 375-378.

De Robertis, A., P. Di Giacomo, & C. Foti 1995. Analitica Chimica Acta, 300; 45.

Herbeli, A. & J. C. Westll 1994. FITEQL: a program for determination of chemical equilibrium constants from experimental data, Report 94-01, Oregon State University.

Jackson, G. E. & L.F. Seymour 1995. Talanta, 42; 5.

Jackson, G. E. & L.F. Seymour 1995. *Talanta*, 42; 9.

- Keizer, M. & W. H. van Riemsdijk 1999. ECOSAT (version 4.5): Equilibrium calculations of speciation and transport; Department of Soil Science, Wagenigen Agricultural University, The Netherlands.
- Kinnburgh, D. G., 1993. FIT user guide, Technical report WD/93/23; British Geological Survey.
- L-Gonzalez, H., J-Reyes, M., R-Hernandez, A.and S-Rios, M., 1997. *Talanta*, 44; 1891.
- Liyanage, J. A. & T. Janaratna 2002. Journal of Chemical Education, 79; 635-638.
- Liyanage, J. A. & C. P. Allas 1999. *Chemical Speciation and Bioavailability*, 11; 103-108.
- Smith, R. M. & A.E. Martell 1989. Critical Stability Constants, Plenum Press, New York.

Back to Contents